

Brogan & Partners

Antitumor Effects of THC

Author(s): James Huff and Po Chan

Source: *Environmental Health Perspectives*, Vol. 108, No. 10 (Oct., 2000), pp. A442-A443

Published by: Brogan & Partners

Stable URL: <http://www.jstor.org/stable/3435034>

Accessed: 23/01/2010 23:50

Your use of the JSTOR archive indicates your acceptance of JSTOR's Terms and Conditions of Use, available at <http://www.jstor.org/page/info/about/policies/terms.jsp>. JSTOR's Terms and Conditions of Use provides, in part, that unless you have obtained prior permission, you may not download an entire issue of a journal or multiple copies of articles, and you may use content in the JSTOR archive only for your personal, non-commercial use.

Please contact the publisher regarding any further use of this work. Publisher contact information may be obtained at <http://www.jstor.org/action/showPublisher?publisherCode=brogpart>.

Each copy of any part of a JSTOR transmission must contain the same copyright notice that appears on the screen or printed page of such transmission.

JSTOR is a not-for-profit service that helps scholars, researchers, and students discover, use, and build upon a wide range of content in a trusted digital archive. We use information technology and tools to increase productivity and facilitate new forms of scholarship. For more information about JSTOR, please contact support@jstor.org.



Brogan & Partners is collaborating with JSTOR to digitize, preserve and extend access to *Environmental Health Perspectives*.

<http://www.jstor.org>

to approximately 50% (4,5). *FU* can vary 15-fold among species, with humans in the middle of the range (12). This variability is an order of magnitude higher than the reproducibility of the same measurement among laboratories (4–6,12). Owing to the lack of human excretion data, fractional excretion values for the rat were used for some congeners. Therefore, our exposure estimates are probably reliable within an order of magnitude.

Exposures for the general population, estimated by the CERHR Phthalates Expert Panel from published data, are in good agreement with our calculated human daily intake estimates based on CDC median values and presented in Table 2. However, the maximal values of excreted monoesters (1) indicate that some individual exposures are substantially higher than previously estimated for the general population.

Women of reproductive age appear to be exposed to higher levels of di-*n*-butyl phthalate than are the remainder of the study population. This is particularly evident in the 95th percentile column for *n*-butyl phthalate in Table 2, where the estimated exposure values for women 20–40 years of age are approximately 5 times greater than the corresponding values for the other 192 individuals in the study.

The data reported by Blount et al. (1) will certainly lead to further efforts to derive accurate estimates of human exposures based on urinary metabolite levels. In addition, their data lead to several questions that should be addressed in the immediate future; for example:

- What are the sources and circumstances of exposure that result in a higher urinary level of diethyl phthalate metabolites than of the other six phthalates studied?
- What is the evidence for reproductive and developmental toxicity of diethyl phthalate?
- What are the sources and circumstances of

exposure that result in some women of reproductive age having higher urinary levels of *n*-butyl phthalate than the remainder of the study population?

- At what levels are humans exposed to other phthalates not included in this study?

It is important that answers to these and related questions be pursued by public health agencies including the NIEHS/NTP.

Michael C. Kohn

Frederick Parham

Scott A. Masten

Christopher J. Portier

Michael D. Shelby

Environmental Toxicology Program

National Institute of Environmental

Health Sciences

Research Triangle Park, North Carolina

E-mail: kohn@niehs.nih.gov

John W. Brock

Larry L. Needham

National Center for Environmental Health

Centers for Disease Control and Prevention

Atlanta, Georgia

REFERENCES AND NOTES

- Blount BC, Silva MJ, Caudill SP, Needham LL, Pirkle JL, Sampson EJ, Lucier GW, Jackson RJ, Brock JW. Levels of seven urinary phthalate metabolites in a human reference population. *Environ Health Perspect* 108:979–982 (2000).
- Tanaka A, Matsumoto A, Yamah T. Biochemical studies on phthalic esters. III. Metabolism of dibutyl phthalate (DBP) in animals. *Toxicology* 9:109–123 (1978).
- Foster PMD, Cook MW, Thomas LV, Walters DG, Gangolli SD. Differences in urinary metabolic profile from di-*n*-butyl phthalate-treated rats and hamsters. A possible explanation for species differences in susceptibility to testicular atrophy. *Drug Metab. Dispos* 11:59–61 (1983).
- Nativelle C, Picard K, Valentin I, Lhuguenot JC, Chagnon MC. Metabolism of *n*-butyl benzyl phthalate in the female Wistar rat. Identification of new metabolites. *Food Chem Toxicol* 37:905–917 (1999).
- Eigenberg DA, Bozigan HP, Carter DE. Distribution, excretion, and metabolism of butylbenzyl phthalate in the rat. *J Toxicol Environ Health* 17:445–456 (1986).
- Kluwe WM. Overview of phthalate ester pharmacokinetics in mammalian species. *Environ Health Perspect* 45:3–9 (1982).
- Peck CC, Albro PW. Toxic potential of the plasticizer di(2-ethylhexyl) phthalate in the context of its disposition and metabolism in primates and man. *Environ Health Perspect* 45:11–17 (1982).
- Albro PW, Moore B. Identification of the metabolites of simple phthalate diesters in rat urine. *J Chromatogr* 94:209–218 (1974).
- Castle L. Personal communication.
- Harper HA, Rodwell VW, Mayes PA. Review of Physiological Chemistry. Los Altos, CA: Lange Medical Publications, 1977.
- The National Toxicology Program (NTP) Center for the Evaluation of Risks to Human Reproduction (CERHR). Available: <http://cerhr.niehs.nih.gov> [cited 14 September 2000].
- Albro PW, Corbett JT, Schroeder JL, Jordan S, Matthews HB. Pharmacokinetics, interactions with macromolecules and species differences in metabolism of DEHP. *Environ Health Perspect* 45:19–25 (1982).

Antitumor Effects of THC

1-Trans-delta⁹-tetrahydrocannabinol (THC), the main active component of marijuana, has been shown to exhibit anticancer activity (1,2). Galve-Roperch et al. (1) reported that intratumoral administration of THC induces apoptosis of transformed neural cells in culture, and also induces a considerable regression of malignant gliomas in Wistar rats and in mice deficient in recombination activating gene 2. These authors suggest that their "results may provide the basis for a new therapeutic approach for the treatment of malignant gliomas." Regarding this interesting finding, we believe it is important to highlight the previous National Toxicology Program's long-term *in vivo* bioassay results that showed definite antitumor activity of THC (3,4). This also appears relevant to the current controversy, at least in the United States, regarding the use of marijuana in clinical medicine.

Experimentally, groups of 60–70 male and female rats were administered 0, 12.5, 25, or 50 mg THC/kg body weight (bw), and male and female mice were given 0, 125, 250, or 500 mg THC/kg bw in corn oil by gavage for 104–106 weeks (3,4). During this 2-year period, individual animal body weights were reduced compared to controls, although all groups consumed the same amounts of food. More importantly, survival in all THC groups of male and female rats was significantly greater than the controls. For mice, survival was comparable among groups except for the high-dose males. Clinical findings in the THC groups included lethargy followed by hyperactivity, convulsions, and seizures, which occurred typically during and immediately after dosing or handling.

In both rats and mice, no increased incidences of neoplasms were considered related to the administration of THC (3,4). In fact, for several organ systems the incidences of background tumors in these strains were actually reduced. The incidences of mammary gland fibroadenomas and uterine stromal

Table 3. Comparison of estimated exposures ($\mu\text{g}/\text{kg}/\text{day}$) to 97 women aged 20–40 years to the rest of the population (192 individuals)^a based on extrapolated intake from urinary metabolites (Equation 1) measured by Blount et al. (1).

Monoester	Diester	Minimum	Median	95th percentile	Maximum
Ethyl	Diethyl	0.90	13	90	170
		< LOD	11	130	320
<i>n</i> -Butyl	Di- <i>n</i> -butyl	0.24	1.7	32	113
		0.084	1.4	6.5	50
Benzyl	<i>n</i> -Butyl benzyl	0.094	1.2	4.5	7.8
		0.11	0.78	3.4	29
Cyclohexyl	Dicyclohexyl	< LOD	0.051	0.24	0.45
			0.012	0.25	2.3
2-Ethylhexyl	Di(2-ethylhexyl)	< LOD	0.71	3.8	10
			0.71	3.5	46
<i>n</i> -Octyl	Di- <i>n</i> -octyl	< LOD	< LOD	0.65	1.5
			0.015	1.0	13
<i>i</i> -Nonyl	Di- <i>i</i> -nonyl	< LOD	< LOD	3.7	7.8
				1.4	22

^aValues for women aged 20–40 years in boldface; remaining values are for the rest of population.

polyps were decreased in THC groups of female rats, as were incidences of pituitary gland adenomas, interstitial cell adenomas of the testis, and pancreatic adenomas in THC-treated male rats. Concerning nonneoplastic lesions in mice, increases of thyroid gland follicular cell hyperplasia occurred in all THC groups, and increases of forestomach hyperplasia and ulcers occurred in THC groups of male mice; yet, no THC-related tumors were observed to progress from these toxic lesions. This common lack of correlation between toxicity and carcinogenicity has long been known (5–7). Regarding carcinogenic activity of THC in mice, thyroid gland follicular cell adenomas were somewhat increased only in the lowest THC-dosed group of mice (125 mg/kg); thyroid gland follicular cell adenomas were found in 0/62 control males versus 6/60, 3/61, and 1/57 mice treated with 125, 250, and 500 mg THC/kg bw, respectively, and in 4/60 control females versus 9/60, 3/60, and 1/60 mice); these were considered not significantly related to THC (3,4). However, there were significant decreases observed for both benign and malignant liver tumors in male and female mice.

The reduced body weights in these long-term studies may have been contributory to the lowered tumor rates (8–10), as most of the reductions in tumor incidences occurred in hormone-controlled organs (11). This should not detract from the overall antitumor effects of THC observed in both sexes of these species and strains. Until further studies are accomplished, these reductions in tumor incidences in six organs should be considered caused by or associated with administration of THC.

Our 2-year studies (3,4) showed that the observed THC antitumor effects are not confined to the site of injection or administration, and these antitumor effects seem to affect a range of “spontaneous” tumors commonly found in rats and mice. Consequently, the THC-associated antitumor effects are systemically active and are applicable to different tumor types at different organ sites. Again, this lack of specificity might lend credence to the notion that these effects are hormonally mediated and likely related to the observed decreases in body weights. Nonetheless, there were significant reductions in total benign and malignant tumors in all organs combined for both species after THC exposure: in male rats, tumors were found in 98% of controls versus 98, 92, and 90% of groups treated with 12.5, 25, and 50 mg THC/kg bw, respectively; in female rats, tumors were found in 88% of controls versus 82, 86, and 70% of treated groups. Most strikingly, in male mice tumors were found in 73% of controls

versus 55, 44, and 30% of male mice treated with 0, 125, 250, and 500 mg THC/kg bw, respectively, and in female mice, tumors were found in 77% of controls versus 52, 43, and 27% of treated groups (3,4).

Interestingly, the dose levels used by Galve-Roperh et al. (1) were similar to those used in our studies (3,4). Their findings also agreed with ours in that THC administration did not affect either food or water intake or hematologic profiles and general clinical chemistry of the animals. Perhaps further animal bioassay studies should be done to learn more about the antitumor effects of THC. For example, animals could be exposed to known carcinogens (e.g., 9,10-dimethylbenz[*a*]anthracene exposure resulting in mammary gland tumors) to determine if THC would block this carcinogenic activity, or transgenic animals (12,13) could be used in an attempt to better clarify the mechanism(s) of THC anticarcinogenic activity. More definition of dose-response-antitumor activity relations would be useful, as would studies using paired feeding, to better define the influence of reduced body weight on tumor incidences.

With respect to genetic toxicology (4), THC was not mutagenic in *Salmonella typhimurium* strains TA97, TA98, TA100, or TA1535 with or without rat and hamster liver S9 fractions. In cultured Chinese hamster ovary (CHO) cells, THC induced sister chromatid exchanges at the highest dose tested in the presence of S9; at this dose level, cell cycle delay indicative of toxicity was observed. THC did not induce chromosomal aberrations in cultured CHO cells with or without S9 metabolic activation enzymes. *In vivo*, no increase in the frequency of micronucleated erythrocytes was observed in the peripheral blood of male or female mice administered THC by gavage for 13 weeks. Accordingly, THC does not appear to be genotoxic.

Long-term carcinogenesis bioassays have historically and traditionally been used primarily to identify those agents that cause cancer in laboratory animals and hence determine which agents represent a significant cancer risk to humans exposed to these carcinogens (14–16). Conversely, these bioassays can and should also be used to identify potential anticarcinogenic agents (17–19). Thus, in our studies, rats and mice that received THC for 2 years exhibited body weight reductions, enhanced survival rates, and decreased tumor incidences in several sites, mainly organs under hormonal control. These earlier experimental carcinogenesis results on THC (3,4) clearly lend further validity to the notion that cannabinoids may indeed be anticarcinogenic (1,2).

**James Huff
Po Chan**

National Institute of Environmental
Health Sciences
Research Triangle Park, North Carolina
E-mail: huff1@niehs.nih.gov

REFERENCES AND NOTES

- Galve-Roperh I, Sanchez C, Cortes ML, del Pulgar TG, Izquierdo M, Guzman M. Anti-tumoral action of cannabinoids: involvement of sustained ceramide accumulation and extracellular signal-regulated kinase activation. *Nat Med* 6(3):313–319 (2000).
- Piomelli D. Pot of gold for glioma therapy. *Nat Med* 6(3):255–256 (2000).
- Chan PC, Sills RC, Braun AG, Haseman JK, Bucher JR. Toxicity and carcinogenicity of delta 9-tetrahydrocannabinol in Fischer rats and B6C3F1 mice. *Fundam Appl Toxicol* 30(1):109–117 (1996).
- NTP. Toxicology and Carcinogenesis Studies of 1-Trans-Delta⁹-Tetrahydrocannabinol (CAS No. 1972-08-3) in F344 Rats and B6C3F1 Mice (Gavage Studies). TR 446. Research Triangle Park, NC:National Toxicology Program, 1996.
- Hoel DG, Haseman JK, Hogan MD, Huff J, McConnell EE. The impact of toxicity on carcinogenicity studies: implications for risk assessment. *Carcinogenesis* 9(11):2045–2052 (1988).
- Huff J. Absence of morphologic correlation between chemical toxicity and chemical carcinogenesis. *Environ Health Perspect* 101(suppl 5):45–53 (1993).
- Huff J. Chemical toxicity and chemical carcinogenesis. Is there a causal connection? A comparative morphological evaluation of 1500 experiments. *IARC Sci Publ* 116:437–475 (1992).
- Rao GN, Piegorsch WW, Crawford DD, Edmondson J, Haseman JK. Influence of viral infections on body weight, survival, and tumor prevalence of B6C3F1 (C57BL/6N x C3H/HeN) mice in carcinogenicity studies. *Fundam Appl Toxicol* 13(1):156–164 (1989).
- Haseman JK, Young E, Eustis SL, Hailey JR. Body weight-tumor incidence correlations in long-term rodent carcinogenicity studies. *Toxicol Pathol* 25(3):256–263 (1997).
- Haseman JK. The National Toxicology Program experience with dietary restriction: does the manner in which reduced body weight is achieved affect tumor incidence? *Int J Toxicol* 17(suppl 2):119–134 (1998).
- Huff J, Boyd J, Barrett JC, eds. Cellular and molecular mechanisms of hormonal carcinogenesis: environmental influences. *Prog Clin Biol Res* 394: 1–479 (1996).
- Tennant R. Transgenic mouse models in chemical carcinogenesis studies. *Arch Toxicol Suppl* 16:261–270 (1994).
- Tennant RW, French JE, Spalding JW. Identifying chemical carcinogens and assessing potential risk in short-term bioassays using transgenic mouse models. *Environ Health Perspect* 103:942–950 (1995).
- Huff J. Long-term chemical carcinogenesis bioassays predict human cancer hazards. Issues, controversies, and uncertainties. *Ann N Y Acad Sci* 895:56–79 (1999).
- Huff J. Animal and human carcinogens [Letter]. *Environ Health Perspect* 107:A341–A342 (1999).
- Huff J. Value, validity, and historical development of carcinogenesis studies for predicting and confirming carcinogenic risks to humans. In: *Carcinogenicity Testing, Predicting, & Interpreting Chemical Effects* (Kitchin KT, ed). New York:Marcel Dekker, 1999:21–123.
- Douglas JF, Huff J, Peters AC. No evidence of carcinogenicity for L-ascorbic acid (vitamin C) in rodents. *J Toxicol Environ Health* 14(4):605–609 (1984).
- Chhabra RS, Huff JE, Haseman J, Hall A, Baskin G, Cowan M. Inhibition of some spontaneous tumors by 4-hexylresorcinol in F344/N rats and B6C3F1 mice. *Fundam Appl Toxicol* 11(4):685–690 (1988).
- Haseman JK, Johnson FM. Analysis of National Toxicology Program rodent bioassay data for anticarcinogenic effects. *Mutat Res* 350(1):131–141 (1996).